

SYNTHESIS OF OPTICALLY PURE ENKEPHALIN ANALOG, [D-Ala², Leu⁵-ol]ENKEPHALIN,
USING CHIRAL β -LACTAMS AS SYNTHETIC INTERMEDIATES

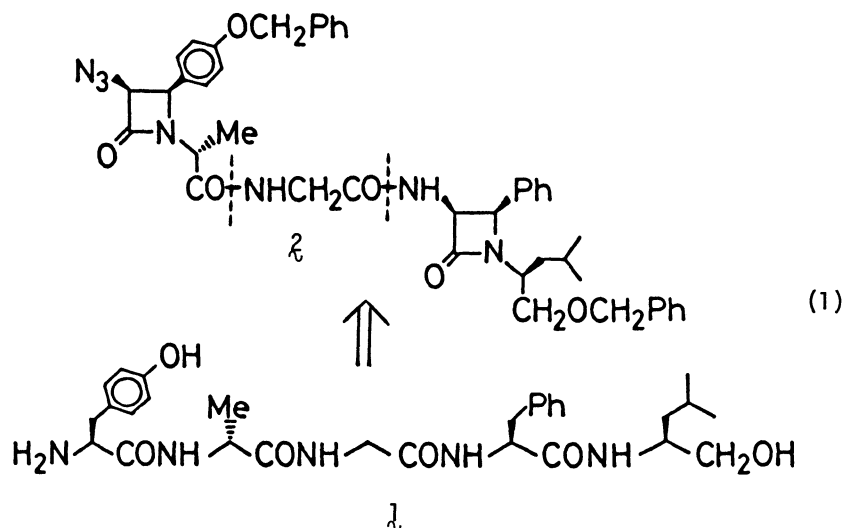
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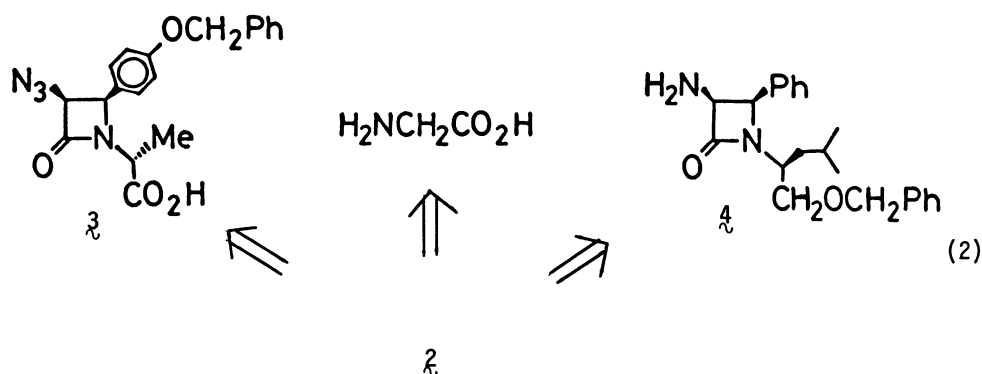
Optically pure pentapeptide, L-Tyr-D-Ala-Gly-L-Phe-L-Leu-ol (**1**) which is an analog of opioid hormone, enkephalin, was successfully synthesized by using β -lactam building blocks as chiral synthons of peptide units.

Recently, we have developed an entirely new method for peptide synthesis via β -lactam intermediates,¹⁻⁵ which is of considerable advantage to the manipulation of peptides because of the high solubility of the β -lactam intermediates in usual organic solvents and a good performance in chromatography on silica gel. In the preceding paper,⁵ we reported a successful application of the β -lactam method to the synthesis of [Leu⁵]enkephalin t-butyl ester which was, however, racemic except leucine residue. Now, we describe here the synthesis of optically pure [D-Ala², Leu⁵-ol]enkephalin (**1**) which is a potent long-lasting analog of enkephalin,⁶ an opioid hormone, by using chiral β -lactam building blocks.

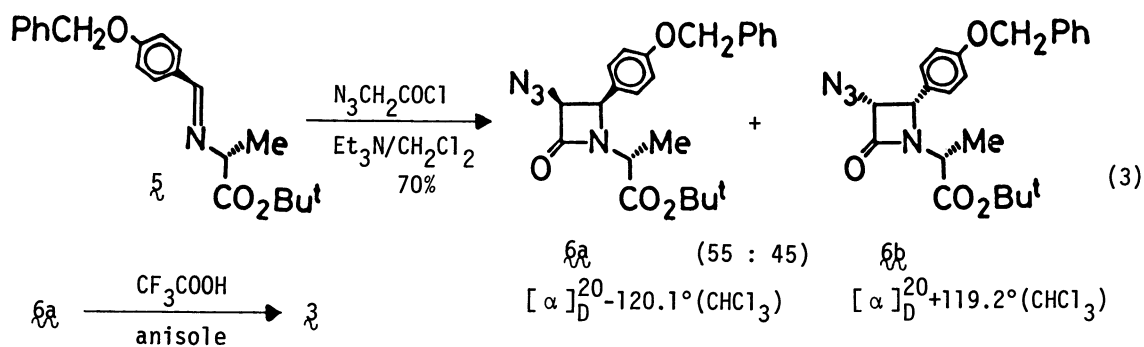
As the final precursor of [D-Ala², Leu⁵-ol]enkephalin (**1**) we planned, based on the retrosynthetic scheme (eq. 1), to synthesize a chiral bis- β -lactam (**2**) which would readily be converted to **1** by hydrogenolysis on palladium.¹⁻⁵



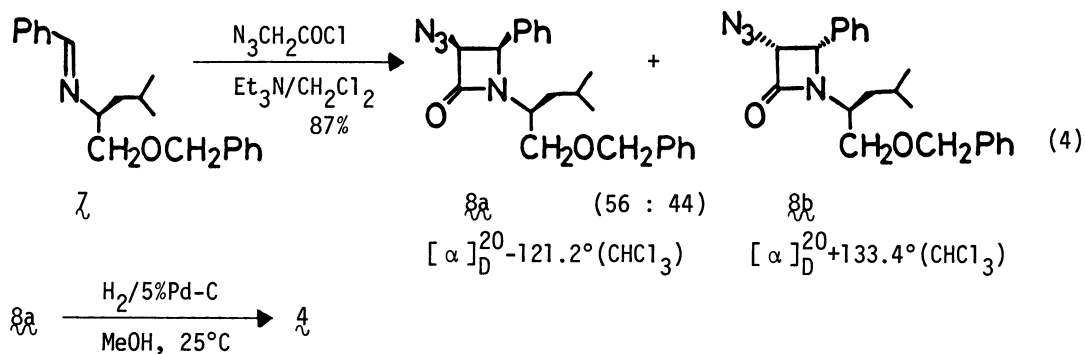
The bis- β -lactam (ζ) consists of three units, i.e., the β -lactam (\mathfrak{z}) which is a synthon of Tyr-(D)-Ala, glycine, and the β -lactam ($\mathfrak{4}$) which is a synthon of Phe-Leu-ol (eq. 2).



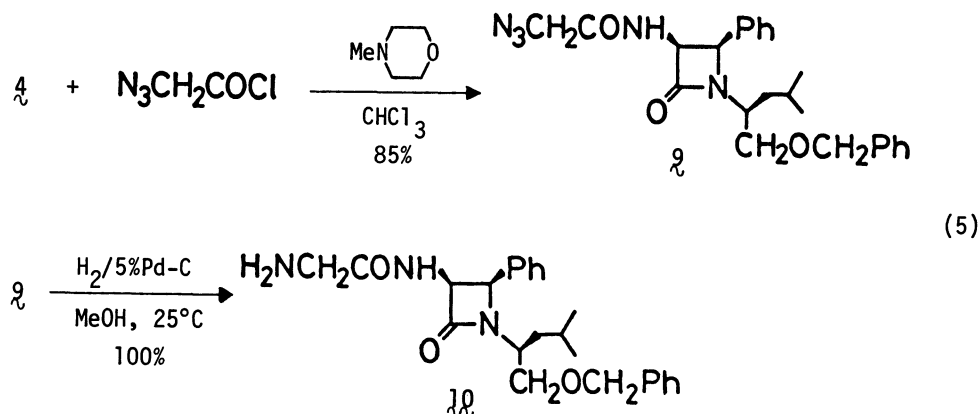
The β -lactam (\mathfrak{z}) was prepared via the in situ azidoketene addition to *t*-butyl 4-benzyloxybenzylidene-(D)-alaninate ($\mathfrak{5}$) followed by HPLC separation of two diastereomers ($\mathfrak{6a}$ and $\mathfrak{6b}$)⁷ on silica gel (n-hexane/AcOEt=2/1) and deprotection of $\mathfrak{6a}$ (eq. 3).



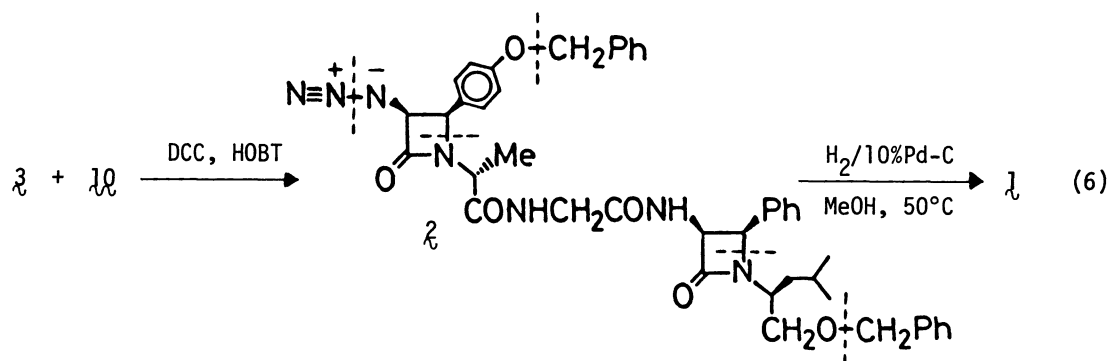
In a similar manner, the β -lactam ($\mathfrak{4}$) was prepared from benzylideneleucinol benzyl ether ($\mathfrak{7}$) and azidoacetyl chloride followed by HPLC separation of two diastereomers ($\mathfrak{8a}$ and $\mathfrak{8b}$)⁸ on silica gel (n-hexane/AcOEt=4/1), and reduction of the azide group of $\mathfrak{8a}$ (eq. 4).



We employed azidoacetyl chloride as glycine synthon. Thus, the β -lactam ($\mathbf{4}$) was azidoacetylated to give the azidoacetyl- β -lactam ($\mathbf{9}$), which was transformed to the tripeptide synthon ($\mathbf{10}$) by the reduction of azide moiety of $\mathbf{9}$ (eq. 5).



The coupling of Tyr-(D)-Ala synthon ($\mathbf{3}$) and Gly-Phe-Leu-ol synthon ($\mathbf{10}$) by using dicyclohexylcarbodiimide (DCC) and 1-hydroxybenzotriazole (HOBT) in dimethylformamide gave the bis- β -lactam ($\mathbf{2}$),⁹ which is the planned final precursor of $\mathbf{1}$, in 84% yield after purification on silica gel column (AcOEt). Then, the pentapeptide synthon ($\mathbf{2}$) was submitted to hydrogenolysis on 10% Pd-C in methanol at 50°C to give $\mathbf{1}^{10}$ in 85% yield through the reductive cleavage of two β -lactam rings and the deprotection of two hydroxy groups and an azide group all at once (eq. 6).



It is noteworthy that the β -lactam ring of $\mathbf{3}$ acts not only as tyrosine synthon but also as excellent protecting group of D-alanine. According to the established rationale of the mechanism of racemization during peptide synthesis, the formation of oxazolone using an acylamino proton or an alkoxy-carbonylamino proton is crucial,¹¹ which is more or less inevitable as far as ordinary protecting groups are employed. However, in the Tyr-(D)-Ala synthon ($\mathbf{3}$), two amino protons of (D)-alanine are protected by the β -lactam ring; the racemization at chiral center cannot take place via oxazolone formation. Actually, no racemization was detected during the DCC-HOBT coupling of $\mathbf{3}$ and $\mathbf{10}$. This must be another advantageous feature of the β -lactam method.

As we have already disclosed, β -lactam building blocks exhibited good solubility toward usual organic solvents, readily purified on usual silica gel column, and fully characterized by spectroscopic analyses. In addition to these, i) the β -lactam building blocks, $6a$ and $6b$, which were not used in the present synthesis, act as the chiral synthons of (D)-Tyr-(D)-Ala and (D)-Phe-Leu-ol, respectively: These are useful for the synthesis of other analogs, and ii) a variety of aromatic substituents, e.g., p-fluorophenyl, 3,4-dihydroxyphenyl, indole, furan, and pyrrole, etc., can be introduced to the β -lactam building blocks simply by using the corresponding aromatic aldehydes. These characteristics together with the stereochemical stability of β -lactam building blocks may provide a powerful device for the synthesis of various physiologically active oligopeptides.

References

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3. N. Hatanaka and I. Ojima, Chem. Lett., 1980, 231.
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6. As for the design of enkephalin analogs having higher potency and longer duration of opioid activity, see J. S. Morley, Ann. Rev. Pharmacol. Toxicol., 20, 81 (1980); C. R. Beddell, R. B. Clark, G. W. Hardy, L. A. Lowe, F. B. Utaba, J. R. Vane, S. Wilkinson, K. J. Chang, P. Cuatrecasas, and R. J. Miller, Proc. Royal Soc. London, 198, 249 (1977), and references therein.
7. The absolute configurations of $6a$ and $6b$ were unambiguously determined on the basis of NMR and HPLC analyses by the comparison of the two diastereomers of Tyr-(D)-Ala- O^t Bu which were obtained by the hydrogenolysis of $6a$ and $6b$ following our method,¹⁻⁵ with the authentically prepared (L)-Tyr-(D)-Ala- O^t Bu.
8. The absolute configurations of $8a$ and $8b$ were unambiguously determined in a manner similar to the case of $6a$ and $6b$ on the basis of NMR and HPLC analyses by using authentically prepared (L)-Phe-(L)-Leu-ol; It was found that $8a$ gave (L)-Phe-(L)-Leu-ol and $8b$ gave (D)-Phe-(L)-Leu-ol by hydrogenolysis.¹⁻⁵
9. Hygroscopic colorless solid; NMR(CDCl₃) δ 0.92 (d, J=5.5Hz, 3H), 0.94 (d, J=5.5Hz, 3H), 1.07 (d, J=7Hz, 3H), 1.14-2.05 (m, 3H), 3.14-3.42 (m, 2H), 3.56 (d, J=6Hz, 2H), 3.80 (m, 1H), 4.31 (q, J=7Hz, 1H), 4.24 (s, 2H), 4.95 (d, J=5Hz, 1H), 5.01 (ABq, J=5Hz, 2H), 5.03 (s, 2H), 5.41 (d of d, J=5Hz, 9Hz, 1H), 6.85-7.50 (m, 21H); IR(KBr) 3320 (ν_{NH}), 2110 (ν_{N_3}), 1750 ($\nu_{C=O}$), 1660 (Amide I), 1540 (Amide II) cm⁻¹; Anal. Calcd. for C₄₃H₄₇N₇O₆·3/2H₂O; C, 65.80; H, 6.42; N, 12.49. Found: C, 65.58; H, 6.23; N, 12.38.; [α]_D²⁰ -55.3° (c=1.07, CHCl₃).
10. Extremely hygroscopic colorless solid; IR(KBr) 3300 (ν_{NH} , ν_{OH}), 1655 (Amide I), 1540 (Amide II) cm⁻¹; Anal. Calcd. for C₂₉H₄₁N₅O₆·5H₂O: C, 53.94; H, 7.90; N, 10.84. Found: C, 53.75; H, 7.47; N, 10.44.; [α]_D²⁰ +15.8° (c=0.91, MeOH). The product was also identified by comparing the HPLC chromatogram with that of authentically prepared HCl·H-(L)-Tyr-(D)-Ala-Gly-(L)-Phe-(L)-Leu-ol. HPLC analysis was carried out using a column packed with TOYO SODA LS 410K (ODS SIL) and MeOH-H₂O.
11. e.g., M. Bodanszky in "The Peptides", Vol. 1, ed. by E. Gross and J. Meienhofer, Academic Press, New York, 1979, pp 152-156.

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